Total Synthesis of Five Cacalol Families at Different Oxidation Stages, Modified Furanoeremophilane Sesquiterpenes from *Cacalia* and *Senecio* Species

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(Received November 14, 2003; CL-031102)

The recent findings of antihyperglycemic and antimicrobial activities for a modified furanoeremophilane sesquiterpene cacalol (1) prompted us to achieve the first total synthesis of five cacalol families (\pm)-2–4, 5, and 6 which might also be expected to indicate such potent biological activities.

Cacalia decomposita A. Gray,¹ a compositae widely distributed in the northern part of Mexico, is a shrub popularly known as matarique and maturin. Matarique is a medicinal plant complex of Mexico which includes perennial herbs with thin, fascicled roots extending from a pubescent root crown, the concoction of which is drunk, alone or in combination with other herbs, for treating diabetes, kidney pain, and rheumatism; it can also be applied as a wash or cataplasm to treat wounds and skin ulcers.² Extensive chromatography of a hexane extract from the roots of Cacalia decomposita A. Gray afforded a modified eremophilane cacalol (1), which has been shown to be the first representative of a new class of compounds (cacalol families) possessing the furotetralin ring system.³ The structures of 1 and related compounds⁴ were the subject of several structural revisions,⁵ and the final structures were confirmed by chemical synthesis⁶ and degradation studies.⁷



Recently, in vivo bioassay-directed fractionation of an extract from the roots of *Cacalia decomposita* A. Gray revealed that **1** exhibits antihyperglycemic⁸ and antimicrobial⁹ activities. Therefore, other known cacalol families, such as 14-oxocacalol methyl ether (**2**),¹⁰ 14-oxo-1,2-dehydrocacalol methyl ether (**3**),^{10,11} 14-hydroxy-1,2-dehydrocacalol methyl ether (**4**),¹¹ and 14-nordehydrocacalohastine (**5**)¹² isolated from *Senecio* species and maturinin (**6**)^{4,11b} isolated from *Cacalia decomposita* A. Gray, might also be expected to possess similar biological activities. We took an interest in biological activities of the furano-eremophilanes **2–6**, the oxidation stages of which were different from those of **1** at the C-ring and C14 position. In this paper, we report the first total synthesis of these cacalol families (±)-**2–4**, **5**, and **6** via stepwise regioselective dehydrogenation of the C-ring.

The retrosynthetic analysis of 2-6 is depicted in Scheme 1. These compounds 2-6 possess a variety of oxidation states and substituents at the C-ring and C6 position, respectively; therefore, we envisaged tetralin 7 as a possible intermediate, because 7 will allow stepwise regioselective dehydrogenation of the Cring and introduction of various substituents at C6. The intermediate 7 will be derived from tetralone 9 via the furan ring formation in 8. 9 will be constructed from commercially available γ -valerolactone 10 and 4-bromoveratrole 11 by Friedel–Crafts reaction.



Scheme 1. Retrosynthetic analysis of cacalol families 2-6.

The intermolecular regioselective Friedel-Crafts alkylation of 11 with 10 gave a mixture 12 of two carboxylic acids, which without separation into each component was successively treated with AcCl in MeOH and subsequent Me₂SO₄ to afford a single methyl ester 13 in 54% yield over 3 steps (Scheme 2).^{6b} Saponification of the ester 13 provided carboxylic acid 14, an intramolecular Friedel-Crafts acylation of which proceeded by $(CF_3CO)_2O$ in $CF_3CO_2H^{13}$ to furnish tetralone 9. Since it was found that selective demethylation of the 8-MeO group in 9 was difficult, we chose to alkylate the 8-OH group with a C₃ unit required for the furan ring formation after removing both methyl groups. Treatment of 9 with BBr₃ yielded catechol 15, etherification of which with chloroacetone could regioselectively be achieved at the 8-OH to give ether 16 in 86% yield. After methylation of 16, the resulting 17 was subjected to reduction with NaBH₄ and CF₃CO₂H to afford monoketone 8 reduced only at the benzylic carbonyl group and further reduced alcohol 18, respectively, both of which were mutually convertible in high yields by a usual manner. Cyclization of 8 with CF3SO3H resulted in the furan derivative 7, which could be dehydrogenated with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ)¹⁴ to provide naphthofuran 19. Debromination of 19 with LiAlH₄ produced 14-nordehydrocacalohastine (5) in a quantitative yield. The spectral characteristics of synthetic 5^{15} were in good agreement with those reported for the natural product.¹²

Next, we turned our attention to syntheses of the other cacalol families **2–4** and **6** with 6-substituents. First, we examined modification at the C6 position in the furotetralin **7**; however, it has been found difficult, probably owing to steric hindrance by the two lateral methyl groups at C4 and C11 positions. Therefore, we adopted a C–C bond formation at C6 before constructing the furan ring. After methoxymethyl (MOM) protection of the alcohol **18**, lithium–halogen exchange in the resultant bromide **20** followed by formylation furnished aldehyde **21** in 90% yield (Scheme 3). Deprotection of the MOM ether in **21** and subsequent Swern oxidation¹⁶ of the alcohol **22** yielded ke-



Scheme 2. Reaction conditions: a) 10, AlCl₃, (CHCl₂)₂, 70 °C; b) AcCl, MeOH, rt, 14 h; c) Me₂SO₄, K₂CO₃, acetone, reflux, 46 h, 54% (3 steps); d) NaOH, MeOH, 40 °C, 24 h, 97%; e) (CF₃CO)₂O, CF₃CO₂H, 40 °C, 3 h, 64%; f) BBr₃, CH₂Cl₂, -78 to -40 °C, 18 h, 98%; g) chloroacetone, Li₂CO₃, 70 °C, 17 h, 86%; h) Me₂SO₄, K₂CO₃, acetone, reflux, 14 h, 77%; i) NaBH₄, CF₃CO₂H, 0 °C to rt, 4 h; j) NaBH₄, MeOH, rt, 14 h, 96%; k) Jones oxidation, 92%; l) CF₃SO₃H, CH₂Cl₂, 0 °C, 30 min, 57%; m) DDQ, CH₂Cl₂, 0 °C, 5 h, 41%; n) LiAlH₄, THF, 0 °C, 4 h, 100%.



Scheme 3. Reaction conditions: a) MOMCl, *i*-Pr₂NEt, CH₂Cl₂, 97%; b) *n*-BuLi, THF, -78 °C, then DMF, 1 h, 90%; c) 3 N HCl, THF, 40 °C, 100%; d) Swern oxidation, 84%; e) CF₃SO₃H, CH₂Cl₂, 0 °C to rt, 5 h, 99%; f) DDQ, benzene, 0 °C to rt, 3 h; g) Burgess reagent, benzene, rt, 37%; h) *o*-chloranil, CH₂Cl₂, rt, 20 h, 54%; i) LiAlH₄, THF, 0 °C, 5 min, 69%.

tone **23**, which was cyclized with CF₃SO₃H to give (±)-14-oxocacalol methyl ether (**2**) in 99% yield. The spectral characteristics of synthetic (±)-**2** were also consistent with those reported for the natural product.¹⁰ After many experimentations, oxidation of **2** employing 1.85 equiv. of DDQ afforded regioselectively 1,2-dehydrogenated (±)-14-oxo-1,2-dehydrocacalol methyl ether (**3**)^{11a} and maturinin (**6**),⁴ respectively, along with 1-hydroxylated compound **24**, which could be transformed into **3** by dehydration with Burgess reagent.¹⁷ **6** was also able to be derived from (±)-**3** by dehydrogenation with *o*-chloranil. LiAlH₄ reduction of **3** gave (±)-14-hydroxy-1,2-dehydrocacalol methyl ether (**4**)^{11a} as well.

In conclusion, we have accomplished the first total synthesis of five cacalol families (\pm) -2–4, 5, and 6, which might be expected to exhibit such antihyperglycemic and antimicrobial activities as those of cacalol (1). Bioassay for synthetic 2–6 and ap-

plication of this synthetic strategy to many other cacalol families are in progress.

M. Doe thanks Osaka City University for the OCU Grant for Graduate Course Students.

References and Notes

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- 15 **5**: $\delta_{\rm H}$ (300 MHz, CDCl₃) 8.21 (1H, d, J = 8.4 Hz), 7.72 (1H, s), 7.47 (1H, q, J = 1.3 Hz), 7.33 (1H, dd, J = 8.4, 6.8 Hz), 7.27 (1H, d, d)J = 6.6 Hz), 4.35 (3H, s), 2.75 (3H, s), 2.33 (3H, d, J = 1.3 Hz); $\delta_{\rm C}$ (75 MHz, CDCl₃) 143.1, 142.3, 138.5, 133.8, 131.8, 130.4, 125.0, 124.5, 123.8, 120.3, 115.7, 107.1, 60.8, 20.3, 8.0. (\pm)-2: $\delta_{\rm H}$ $(400 \text{ MHz}, \text{ CDCl}_3)$ 10.68 (1H, s), 7.42 (1H, q, J = 1.0 Hz), 4.28 (3H, s), 4.03 (1H, qt, J = 7.0, 3.5 Hz), 2.97 (1H, ddd, J = 18.0, 5.9, 2.0 Hz), 2.59 (1H, ddd, J = 18.1, 11.0, 7.2 Hz), 2.37 (3H, d, J = 1.2 Hz), 1.95–1.71 (4H, m), 1.27 (3H, d, J = 7.1 Hz); δ_{C} (100 MHz, CDCl₃) 190.4, 146.6, 144.4, 143.8, 143.2, 131.4, 123.5, 121.8, 116.0, 60.1, 29.3, 28.7, 23.7, 23.5, 16.5, 12.8. (\pm) -3: $\delta_{\rm H}$ $(300 \text{ MHz}, \text{ CDCl}_3)$ 10.70 (1H, s), 7.47 (1H, q, J = 1.1 Hz), 6.90 (1H, dd, J = 9.9, 3.1 Hz), 6.00 (1H, dddd, J = 9.7, 6.5, 2.4, 0.6 Hz), 4.27 (3H, s), 4.09 (1H, quintet, J = 6.8 Hz), 2.51 (1H, ddt, J = 17.3, 6.7, 2.9 Hz), 2.38 (3H, d, J = 1.3 Hz), 2.22 (1H, ddd, J = 17.3, 6.5, 1.5 Hz, 1.17 (3H, d, J = 7.1 Hz); δ_{C} (75 MHz, CDCl₃) 190.4, 145.1, 144.21, 144.18, 141.7, 131.4, 126.1, 121.7, 120.5, 120.1, 116.3, 60.6, 29.9, 27.3, 20.3, 12.7. **6**: $\delta_{\rm H}$ (400 MHz, CDCl₃) 11.08 (1H, s), 8.28 (1H, t, J = 4.9 Hz), 7.56 (1H, q, J = 1.2 Hz, 7.40 (2H, d, J = 5.9 Hz), 4.43 (3H, s), 2.69 (3H, s), 2.33 (3H, d, J = 1.2 Hz). (±)-4: $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.36 (1H, q, J = 1.2 Hz), 6.93 (1H, dd, J = 9.8, 3.2 Hz), 5.94 (1H, dddd, J = 9.8, 6.4, 2.4, 0.9 Hz), 4.92 (2H, s), 4.11 (3H, s), 3.43 (1H, quintet, J = 6.9 Hz), 2.55 (1H, ddt, J = 17.1, 6.3, 2.9 Hz), 2.42 (3H, d, J = 1.2 Hz), 2.24 (1H, ddd, J = 17.1, 6.3, 1.2 Hz), 1.46 (1H, br s), 1.14 (3H, d, J = 7.1 Hz); δ_{C} (100 MHz, CDCl₃) 145.8, 142.3, 140.5, 136.0, 128.7, 124.9, 123.7, 121.2, 121.0, 116.2, 60.8, 57.3, 30.6, 27.7, 21.2, 10.2.
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