Total Synthesis of Five Cacalol Families at Different Oxidation Stages, Modified Furanoeremophilane Sesquiterpenes from Cacalia and Senecio Species

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The recent findings of antihyperglycemic and antimicrobial activities for a modified furanoeremophilane sesquiterpene cacalol (1) prompted us to achieve the first total synthesis of five cacalol families (\pm) -2–4, 5, and 6 which might also be expected to indicate such potent biological activities.

Cacalia decomposita A. Gray, $\frac{1}{1}$ a compositae widely distributed in the northern part of Mexico, is a shrub popularly known as matarique and maturin. Matarique is a medicinal plant complex of Mexico which includes perennial herbs with thin, fascicled roots extending from a pubescent root crown, the concoction of which is drunk, alone or in combination with other herbs, for treating diabetes, kidney pain, and rheumatism; it can also be applied as a wash or cataplasm to treat wounds and skin ulcers.² Extensive chromatography of a hexane extract from the roots of Cacalia decomposita A. Gray afforded a modified eremophilane cacalol (1), which has been shown to be the first representative of a new class of compounds (cacalol families) possessing the furotetralin ring system.³ The structures of 1 and related compounds 4 were the subject of several structural revisions,⁵ and the final structures were confirmed by chemical synthesis⁶ and degradation studies.⁷

Recently, in vivo bioassay-directed fractionation of an extract from the roots of Cacalia decomposita A. Gray revealed that 1 exhibits antihyperglycemic δ and antimicrobial⁹ activities. Therefore, other known cacalol families, such as 14-oxocacalol methyl ether (2) ,¹⁰ 14-oxo-1,2-dehydrocacalol methyl ether (3) ,^{10,11} 14-hydroxy-1,2-dehydrocacalol methyl ether (4) ,¹¹ and 14-nordehydrocacalohastine $(5)^{12}$ isolated from Senecio species and maturinin $(6)^{4,11b}$ isolated from *Cacalia decomposita* A. Gray, might also be expected to possess similar biological activities. We took an interest in biological activities of the furanoeremophilanes 2–6, the oxidation stages of which were different from those of 1 at the C-ring and C14 position. In this paper, we report the first total synthesis of these cacalol families (\pm) -2–4, 5, and 6 via stepwise regioselective dehydrogenation of the C-ring.

The retrosynthetic analysis of 2–6 is depicted in Scheme 1. These compounds 2–6 possess a variety of oxidation states and substituents at the C-ring and C6 position, respectively; therefore, we envisaged tetralin 7 as a possible intermediate, because 7 will allow stepwise regioselective dehydrogenation of the Cring and introduction of various substituents at C6. The intermediate 7 will be derived from tetralone 9 via the furan ring formation in 8. 9 will be constructed from commercially available γ -valerolactone 10 and 4-bromoveratrole 11 by Friedel–Crafts reaction.

Scheme 1. Retrosynthetic analysis of cacalol families 2–6.

The intermolecular regioselective Friedel–Crafts alkylation of 11 with 10 gave a mixture 12 of two carboxylic acids, which without separation into each component was successively treated with AcCl in MeOH and subsequent $Me₂SO₄$ to afford a single methyl ester 13 in 54% yield over 3 steps (Scheme 2).^{6b} Saponification of the ester 13 provided carboxylic acid 14, an intramolecular Friedel–Crafts acylation of which proceeded by $(CF_3CO)_2O$ in $CF_3CO_2H^{13}$ to furnish tetralone 9. Since it was found that selective demethylation of the 8-MeO group in 9 was difficult, we chose to alkylate the 8-OH group with a C_3 unit required for the furan ring formation after removing both methyl groups. Treatment of 9 with BBr₃ yielded catechol 15, etherification of which with chloroacetone could regioselectively be achieved at the 8-OH to give ether 16 in 86% yield. After methylation of 16, the resulting 17 was subjected to reduction with NaBH₄ and CF_3CO_2H to afford monoketone 8 reduced only at the benzylic carbonyl group and further reduced alcohol 18, respectively, both of which were mutually convertible in high yields by a usual manner. Cyclization of 8 with $CF₃SO₃H$ resulted in the furan derivative 7, which could be dehydrogenated with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone $(DDQ)^{14}$ to provide naphthofuran 19. Debromination of 19 with LiAlH₄ produced 14-nordehydrocacalohastine (5) in a quantitative yield. The spectral characteristics of synthetic 5^{15} were in good agreement with those reported for the natural product.¹²

Next, we turned our attention to syntheses of the other cacalol families 2–4 and 6 with 6-substituents. First, we examined modification at the C6 position in the furotetralin 7; however, it has been found difficult, probably owing to steric hindrance by the two lateral methyl groups at C4 and C11 positions. Therefore, we adopted a C–C bond formation at C6 before constructing the furan ring. After methoxymethyl (MOM) protection of the alcohol 18, lithium–halogen exchange in the resultant bromide 20 followed by formylation furnished aldehyde 21 in 90% yield (Scheme 3). Deprotection of the MOM ether in 21 and subsequent Swern oxidation¹⁶ of the alcohol 22 yielded ke-

Scheme 2. Reaction conditions: a) 10, AlCl₃, (CHCl₂)₂, 70 °C; b) AcCl, MeOH, rt, 14 h; c) Me₂SO₄, K₂CO₃, acetone, reflux, 46 h, 54% (3 steps); d) NaOH, MeOH, 40 °C, 24 h, 97%; e) $(CF_3CO)_2O$, CF_3CO_2H , $40°C$, 3h, 64%; f) BBr_3 , CH_2Cl_2 , -78 to -40° C, 18 h, 98%; g) chloroacetone, Li₂CO₃, 70 °C, 17 h, 86%; h) Me₂SO₄, K₂CO₃, acetone, reflux, 14 h, 77%; i) NaBH₄, CF₃CO₂H, 0° C to rt, 4h; j) NaBH₄, MeOH, rt, 14h, 96%; k) Jones oxidation, 92%; l) CF_3SO_3H , CH_2Cl_2 , 0 °C, 30 min, 57%; m) DDQ, CH₂Cl₂, 0 °C, 5h, 41%; n) LiAlH₄, THF, 0°C, 4h, 100%.

Scheme 3. Reaction conditions: a) MOMCl, i -Pr₂NEt, CH₂Cl₂, 97%; b) *n*-BuLi, THF, -78 °C, then DMF, 1 h, 90%; c) 3 N HCl, THF, 40° C, 100% ; d) Swern oxidation, 84% ; e) CF₃SO₃H, CH₂Cl₂, 0° C to rt, 5 h, 99%; f) DDQ, benzene, 0° C to rt, 3 h; g) Burgess reagent, benzene, rt, 37% ; h) o-chloranil, CH_2Cl_2 , rt, 20 h, 54%; i) LiAlH₄, THF, 0 °C, 5 min, 69%.

tone 23, which was cyclized with CF_3SO_3H to give (\pm) -14-oxocacalol methyl ether (2) in 99% yield. The spectral characteristics of synthetic (\pm) -2 were also consistent with those reported for the natural product.¹⁰ After many experimentations, oxidation of 2 employing 1.85 equiv. of DDQ afforded regioselectively 1,2-dehydrogenated (\pm) -14-oxo-1,2-dehydrocacalol methyl ether $(3)^{11a}$ and maturinin (6) , respectively, along with 1-hydroxylated compound 24, which could be transformed into 3 by dehydration with Burgess reagent.¹⁷ 6 was also able to be derived from (\pm) -3 by dehydrogenation with *o*-chloranil. LiAlH₄ reduction of 3 gave (\pm) -14-hydroxy-1,2-dehydrocacalol methyl ether (4) ^{11a} as well.

In conclusion, we have accomplished the first total synthesis of five cacalol families (\pm) -2-4, 5, and 6, which might be expected to exhibit such antihyperglycemic and antimicrobial activities as those of cacalol (1). Bioassay for synthetic 2–6 and application of this synthetic strategy to many other cacalol families are in progress.

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References and Notes

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- 15 5: δ_H (300 MHz, CDCl₃) 8.21 (1H, d, $J = 8.4$ Hz), 7.72 (1H, s), 7.47 (1H, q, $J = 1.3$ Hz), 7.33 (1H, dd, $J = 8.4$, 6.8 Hz), 7.27 (1H, d, $J = 6.6$ Hz), 4.35 (3H, s), 2.75 (3H, s), 2.33 (3H, d, $J = 1.3$ Hz); δ C (75 MHz, CDCl₃) 143.1, 142.3, 138.5, 133.8, 131.8, 130.4, 125.0, 124.5, 123.8, 120.3, 115.7, 107.1, 60.8, 20.3, 8.0. (\pm) -2: δ _H $(400 \text{ MHz}, \text{CDCl}_3)$ 10.68 (1H, s), 7.42 (1H, q, $J = 1.0 \text{ Hz}$), 4.28 (3H, s), 4.03 (1H, qt, $J = 7.0$, 3.5 Hz), 2.97 (1H, ddd, $J = 18.0$, 5.9, 2.0 Hz), 2.59 (1H, ddd, $J = 18.1$, 11.0, 7.2 Hz), 2.37 (3H, d, $J = 1.2$ Hz), 1.95–1.71 (4H, m), 1.27 (3H, d, $J = 7.1$ Hz); δ_c (100 MHz, CDCl3) 190.4, 146.6, 144.4, 143.8, 143.2, 131.4, 123.5, 121.8, 116.0, 60.1, 29.3, 28.7, 23.7, 23.5, 16.5, 12.8. (\pm) -3: δ_H $(300 \text{ MHz}, \text{CDCl}_3)$ 10.70 (1H, s), 7.47 (1H, q, $J = 1.1 \text{ Hz}$), 6.90 $(1H, dd, J = 9.9, 3.1 Hz), 6.00 (1H, dddd, J = 9.7, 6.5, 2.4,$ 0.6 Hz), 4.27 (3H, s), 4.09 (1H, quintet, $J = 6.8$ Hz), 2.51 (1H, ddt, $J = 17.3, 6.7, 2.9$ Hz), 2.38 (3H, d, $J = 1.3$ Hz), 2.22 (1H, ddd, $J = 17.3, 6.5, 1.5 \text{ Hz}$, 1.17 (3H, d, $J = 7.1 \text{ Hz}$); δ_C (75 MHz, CDCl3) 190.4, 145.1, 144.21, 144.18, 141.7, 131.4, 126.1, 121.7, 120.5, 120.1, 116.3, 60.6, 29.9, 27.3, 20.3, 12.7. 6: δ_H (400 MHz, CDCl₃) 11.08 (1H, s), 8.28 (1H, t, $J = 4.9$ Hz), 7.56 (1H, q, $J = 1.2$ Hz), 7.40 (2H, d, $J = 5.9$ Hz), 4.43 (3H, s), 2.69 (3H, s), 2.33 (3H, d, $J = 1.2$ Hz). (\pm)-4: δ _H (400 MHz, CDCl₃) 7.36 (1H, q, $J = 1.2$ Hz), 6.93 (1H, dd, $J = 9.8$, 3.2 Hz), 5.94 (1H, dddd, $J = 9.8, 6.4, 2.4, 0.9$ Hz), 4.92 (2H, s), 4.11 (3H, s), 3.43 (1H, quintet, $J = 6.9$ Hz), 2.55 (1H, ddt, $J = 17.1$, 6.3, 2.9 Hz), 2.42 (3H, d, $J = 1.2$ Hz), 2.24 (1H, ddd, $J = 17.1$, 6.3, 1.2 Hz), 1.46 (1H, br s), 1.14 (3H, d, $J = 7.1$ Hz); δ_C (100 MHz, CDCl₃) 145.8, 142.3, 140.5, 136.0, 128.7, 124.9, 123.7, 121.2, 121.0, 116.2, 60.8, 57.3, 30.6, 27.7, 21.2, 10.2.
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